

## In-vitro antimicrobial activity of *Cymbopogon citratus* Stem extracts

Phang Siao Ze<sup>1</sup>, Chao Xu Yu<sup>1</sup>, Lee Sian Jo<sup>1</sup>, Vetrivel Subramaniyan<sup>2</sup>, Pradeep Kumar Sharma<sup>3</sup>, Dhanalekshmi Unnikrishnan Meenakshi<sup>4</sup>, Velmurugan Chinnasamy<sup>5</sup>, Saraswathy Malayappan Palanisamy<sup>6</sup>, Narra Kishore<sup>7</sup>, Rajasekaran S.<sup>8</sup>, Sanjana Adinarayana<sup>9</sup>, Dinesh Kumar Yadav<sup>10</sup>, Lalit Parihar<sup>11</sup>, Shekhar Prakash Kushwaha<sup>3</sup>, T. Muthuramu<sup>12</sup>, Shivkanya Fuloria<sup>1\*</sup>, Neeraj Kumar Fuloria<sup>1\*</sup>

<sup>1</sup> Faculty of Pharmacy, AIMST University, Bedong, Kedah 08100 Malaysia.

<sup>2</sup> Faculty of Medicine, Bioscience and Nursing, MAHSA University, Kuala Lumpur 42610 Malaysia

<sup>3</sup> Accurate College of Pharmacy, Knowledge Park-III, Greater Noida, U.P., India.

<sup>4</sup> College of Pharmacy, National University of Science and Technology, Postal Code: 130, Muscat, Sultanate of Oman.

<sup>5</sup> Department of Pharmacology, PGP College of Pharmaceutical Science and Research Institute, Namakkal, India.

<sup>6</sup> Department of Microbiology, ESIC Medical College and PGIMS, K K Nagar, Chennai 600078, Tamil Nadu, India.

<sup>7</sup> Department of Pharmaceutical Technology, University College of Engineering, Bharathidasan, Institute of Technology Campus, Anna University, Tiruchirapalli 620024 India.

<sup>8</sup> Department of Pharmacology, College of health and medical sciences, Haramaya University, Harar, Ethiopia

<sup>9</sup> Yenepoya Pharmacy college and research centre, Yenepoya Deemed to be a university, Mangalore-575018 India.

<sup>10</sup> SGT College of Pharmacy, SGT University, Gurugram Haryana 122505, India.

<sup>11</sup> R.V. Northland Institute, Dadri, Greater Noida, U.P., India

<sup>12</sup> Arba Minch University College of Natural Sciences Arba Minch University, Ethiopia.

### Correspondence Address\*

Dr. Shivkanya Fuloria, shivkanya\_fuloria@aimst.edu.my

Dr. Neeraj Kumar Fuloria, neerajkumar@aimst.edu.my

### Abstract

Essential oils have always been a field of interest attributed to their strong antibacterial properties. Present study was intended to determine the in-vitro antimicrobial and antioxidant activities of *Cymbopogon citratus* stems. Study involved successive extraction of *Cymbopogon citratus* stems by Soxhlet extraction using different solvents such as dichloromethane, acetone, ethanol and methanol in increasing order of polarity to yield different solvents extracts. The extracts were investigated for their antimicrobial potential. Among all extracts, dichloromethane extract exhibited highest antimicrobial activity followed by ethanolic extract, acetone extract and lastly methanolic extract. Present study concludes that dichloromethane extracts of *Cymbopogon citratus* stem possess highest inhibitory action over the growth of bacterial strains of *Staphylococcus aureus*, *Streptococcus pyogenes*, *Bacillus cereus*, *Escherichia coli*, *Acinetobacter baumannii* and *Neisseria gonorrhoeae*. The high antimicrobial potential of dichloromethane extract of *Cymbopogon citratus* stem may be due to presence flavonoidal constituents.

**Keywords:** Antimicrobials, *Cymbopogon citratus*, bacteria, stem, extract

### 1. Introduction

The human body microbiota is known to possess a ratio of 1:1 human cells and bacteria [1]. A slight disturbance in this ratio manifest in various infections and diseases [2,3]. The development of microbial resistance and patient tolerance have gradually reduced the

efficacy of commercially available antibiotics on one hand and increased the demand for alternative antibiotics [4,5]. Essential oils have always been a field of interest attributed to their high medicinal value in humans and animals [6,7]. Plants are known to possess high medicinal value [8-18]. Medicinal plants are the good source of natural antimicrobial agents attributed to their defence mechanism against microorganism [19-25]. *Cymbopogon citratus* is a frost-tender clumping perennial grass that is popularly used as a lemony flavouring in Thai, Vietnamese, Laotian and Cambodian cooking and is widely cultivated in Southeast Asia. *C. citratus* is believed to be a native of Malaysia [24]. In English, the herb is commonly known as lemongrass, citronella grass or fever grass. More recently, the aromatic oils of the plant have been extracted and exported for use in perfumes [23]. Lemon grass is common and widespread within its natural range, and also occurs in cultivation. The leaves of *Cymbopogon citratus* have been used in traditional medicine and are often found in herbal supplements and teas. Many effects have been attributed to both their oral consumption and topical use, with modern research supporting many of their alleged benefits. Laboratory studies have shown cytoprotective, antioxidant, and anti-inflammatory properties in vitro as well as antifungal properties [24]. A total of 16 chemical constituents accounting for 93.69 % of the oil, were identified in *C. citratus* among which, geranial (27.04 %), neral (19.93 %) and myrcene (27.04 %) were the major constituents. Lemongrass should be harvested at the appropriate level of maturity in order to achieve high quality essential oil and low production cost. Only 13 compounds were present at each of the maturity stage. Among 13 compounds, only 7 compounds ( $\beta$ -myrcene, 3-undecyne, neral, geranial, nerol, geranyl acetate and juniper camphor) had a concentration of greater than 1% [25]. Development of bacterial resistance to conventional antibiotics necessitates the search for new antibacterial agents [26-31]. Hence the present study was carried out to find out the antibacterial activity of lemongrass oil against the selected pathogenic bacteria.

## 2. Material and methods

The *Cymbopogon citratus* were collected from Laguna Merbok in Sungai Petani, district of Kedah state of Malaysia. The collected plant stems were dried using the oven at 40°C for three consecutive days. The chemical and reagents used in the study were of laboratory grade and were procured from Merck, SD fine, and Sigma Aldrich. For example: dichloromethane, acetone, ethanol, methanol were used for extraction of *Cymbopogon citratus*. Muller Hinton Agar and Nutrient Broth were used as media and Gentamicin as standard for antimicrobial test.

### 2.1. Authentication of plant materials (Preparation of herbarium)

The specimen including stems was collected from *Cymbopogon citratus* plant. The specimen was washed with water and rinsed with 70% ethanol in order to remove dirt particles. The specimen was kept between the newspapers and which was then pressed under the wooden press. The specimen was kept air-dried for one month. The dried specimen was mounted onto an acid-free paper. The herbarium was labeled with its common name, geographical situation, place and date of collection, and other features of plant [32]. The herbarium was then submitted to the Pharmaceutical Chemistry Unit, Faculty of Pharmacy, AIMST University. (Herbarium voucher specimen Accession number AIMST/FOP/15 *Cymbopogon citratus*)

### 2.2. Collection and drying of *Cymbopogon Citratus* stems

The *Cymbopogon citratus* stems were collected, cleaned with distilled water and dried using oven at 40°C for three days. The dried stems were then crushed into coarse powder using

blender. Later on, the obtained powder was subjected to different solvent extraction using Soxhlet apparatus [33-44].

### 2.3. Preparation of *Citrus citratus* stem extracts

The *Cymbopogon citratus* dried stems powder was subjected to successive extraction with dichloromethane, acetone, ethanol and methanol in an increasing order of polarity. About 15 g *Cymbopogon citratus* dried stem powder was placed into the Soxhlet extraction tube. The Soxhlet apparatus was set up and made ready for extraction. Extra care was needed while setting up the apparatus. A few pieces of porcelain chips were placed to reduce the bumping during extraction. The dried stems powder was successively extracted using different solvents such as dichloromethane, acetone, ethanol and methanol. The reaction was started once the solvent was heated for 25 siphonic cycles. After about 25 siphonic cycles (approximately for 8 hours), a change in the colour of the extraction solvent from yellowish/green to colourless was observed in the Soxhlet extracting tube. Lastly, the solution was collected and evaporated using evaporating dish to obtain a concentrated pure extract [45-51]. The experiment was repeated by using different extraction solvent successively in their order of increasing polarity such as: dichloromethane, acetone, ethanol, followed by methanol.

### 2.4. Determination of Antibacterial potential

#### 2.4.1. Preparation of bacterial culture

Six bacterial strains such as: *Staphylococcus aureus*, *Staphylococcus pyogenes*, *Bacillus cereus*: Gram positive bacteria, *Escherichia coli*, *Acinetobacter baumannii*, and *Neisseria gonorrhoeae* were used for the antimicrobial study. The microorganism stock cultures were maintained and stored at 4°C in the refrigerator. The Nutrient Broth and apparatus used in the transfer of microorganisms were previously autoclaved to ensure absence of other organisms. Subcultures were prepared by transferring a loopful of colonies of microorganisms from the stock culture into Nutrient Broth and incubated at 37°C for 24 hours in the shaking incubator. Turbidity of the broth indicated the growth of microorganism [52-60].

#### 2.4.2. Agar Well Diffusion method

Antibacterial potential of the extracts of *Cymbopogon citratus* stems was determined using well diffusion method. Mueller Hinton agar (MHA) medium was used in this test. Approximately 100 µl bacteria suspension was pipetted on the MHA plate and spreaded over the agar surface evenly using a sterile L-shaped glass rod. Study involved gentamicin (0.1 mg/ml), dichloromethane extract (1 mg/ml), acetone extract (1 mg/ml), ethanol extract (1 mg/ml), and methanol extract (1 mg/ml). Holes with diameter of 8-10 mm were punched aseptically with a sterile cork borer on the agar containing bacteria suspension. Volume of approximately 100-200 µl of the gentamicin solution, extracts and phosphate buffer at desired concentration were introduced into the well. The agar plates were incubated at 37°C for 24 hours. The diameters of the zones of inhibition with different extracts were measured in mm. Each test was performed in triplicate and the mean values were recorded [61-68].

#### 2.4.3. Agar Disc Diffusion method

Antibacterial potential of the extracts of *Cymbopogon citratus* stems was determined using disc diffusion method. Mueller Hinton agar (MHA) medium was used in this test. Approximately 100 µl bacteria suspension was pipetted on the MHA plate and spreaded over

the agar surface evenly using a sterile L-shaped glass rod. Study involved gentamicin (0.1 mg/ml), dichloromethane extract (1 mg/ml), acetone extract (1 mg/ml), ethanol extract (1 mg/ml), and methanol extract (1 mg/ml). Dried and sterilized filter paper discs with 6 mm diameter were suspended into the gentamicin solution and extracts solution with the help of forceps. Discs with absorbed extracts were placed on the surface of the agar containing bacteria suspension with sterile forceps. The plates were incubated at 37°C for 24 hours. The diameters of the zones of inhibition by discs with different extracts were measured in mm. Each test was performed in triplicate and the mean values were recorded [69-78].

### 3. Results

#### 3.1. Determination of antimicrobial potential

In present study, the prepared extracts of lemon grass (*Cymbopogon citratus*) stem were evaluated for their antimicrobial potential against various bacterial strains such as *Staphylococcus aureus*, *Streptococcus pyogenes*, *Bacillus cereus*, *Escherichia coli*, *Acinetobacter baumannii* and *Neisseria gonorrhoeae*, using Agar well diffusion and disc diffusion methods for measurement of zone of inhibition. The prepared extracts of lemon grass (*Cymbopogon citratus*) stem were evaluated for their antimicrobial potential against various bacterial strains using well diffusion and disc diffusion methods. The results so obtained are given in table 1 and table 2.

Table 1: Zone of inhibition of the lemon grass (*Cymbopogon Citratus*) extracts using well diffusion method

Microorganism	Component	Zone of inhibition (mm)			
		Reading 1	Reading 2	Reading 3	Average value
<i>Staphylococcus aureus</i>	Positive control <i>Gentamicin (1mg/ml)</i>	16	20	14	16.7
	Dichloromethane	-	-	-	-
	Acetone	-	-	-	-
	Ethanol	-	2	-	-
	Methanol	-	-	-	-
	Negative control	-	-	-	-
<i>Streptococcus pyogenes</i>	Positive control <i>Gentamicin (1mg/ml)</i>	18	15	13	15.3
	Dichloromethane	8	-	4	6
	Acetone	-	3	3	3
	Ethanol	-	6	2	4
	Methanol	-	-	3	-
	Negative control	-	-	-	-
<i>Bacillus cereus</i>	Positive control <i>Gentamicin (1mg/ml)</i>	19	16	14	16.3
	Dichloromethane	8	5	10	7.67
	Acetone	-	3	2	2.5
	Ethanol	-	3	-	-
	Methanol	2	1	2	1.67
	Negative control	-	-	-	-
<i>Escherichia coli</i>	Positive control <i>Gentamicin (1mg/ml)</i>	18	18	15	17
	Dichloromethane	19	15	-	17
	Acetone	-	10	2	6
	Ethanol	-	8	2	5
	Methanol	-	2	-	-
	Negative control	-	-	-	-
<i>Acinetobacter baumannii</i>	Positive control <i>Gentamicin (1mg/ml)</i>	14	19	15	16

	Dichloromethane	6	8	-	7
	Acetone	2	8	2	4
	Ethanol	-	3	-	-
	Methanol	2	-	2	2
	Negative control	-	-	-	-
<i>Neisseria gonorrhoeae</i>	Positive control <i>Gentamicin (1mg/ml)</i>	17	18	13	16
	Dichloromethane	14	13	-	13.5
	Acetone	-	9	7	8
	Ethanol	-	10	-	-
	Methanol	-	2	3	2.5
	Negative control	-	-	-	-

Table 2: Zone of inhibition of the lemon grass (*Cymbopogon Citratus*) extracts using disc diffusion method

Microorganism	Component	Zone of inhibition (mm)		
		Reading 1	Reading 2	Average value
<i>Staphylococcus aureus</i>	Positive control <i>Gentamicin (1mg/ml)</i>	9	10	9.5
	Dichloromethane	-	-	-
	Acetone	-	-	-
	Ethanol	-	1	-
	Methanol	2	3	2.5
	Negative control	-	-	-
<i>Streptococcus pyogenes</i>	Positive control <i>Gentamicin (1mg/ml)</i>	15	8	11.5
	Dichloromethane	-	-	-
	Acetone	3	-	-
	Ethanol	3	8	5.5
	Methanol	4	3	3.5
	Negative control	-	-	-
<i>Bacillus cereus</i>	Positive control <i>Gentamicin (1mg/ml)</i>	18	15	16.5
	Dichloromethane	-	-	-
	Acetone	7	4	5.5
	Ethanol	7	2	4.5
	Methanol	4	3	3.5
	Negative control	-	-	-
<i>Escherichia coli</i>	Positive control <i>Gentamicin (1mg/ml)</i>	17	23	20
	Dichloromethane	-	-	-
	Acetone	6	2	4
	Ethanol	7	11	9
	Methanol	5	4	4.5
	Negative control	-	-	-
<i>Acinetobacter baumannii</i>	Positive control <i>Gentamicin (1mg/ml)</i>	16	18	17
	Dichloromethane	-	-	-
	Acetone	7	-	-
	Ethanol	8	2	5
	Methanol	2	2	2
	Negative control	-	-	-
<i>Neisseria gonorrhoeae</i>	Positive control <i>Gentamicin (1mg/ml)</i>	18	16	17
	Dichloromethane	2	-	-
	Acetone	10	2	6
	Ethanol	10	3	6.5
	Methanol	3	2	2.5
	Negative control	-	-	-

#### 4. Discussion

There is an increasing demand on evaluation of alternative antimicrobial due to development of resistant to currently available antimicrobial. As per the literature survey, *Cymbopogon citratus* is reported as a herbal medicine that is useful in treating various diseases and it is majorly due to its strong antimicrobial activity. Therefore, the investigators planned to work over In-vitro antimicrobial activity of *Cymbopogon citratus* stem extracts. Antimicrobial study was conducted as per standard reference [11,15, 16]. The antimicrobial results of the present study revealed dichloromethane extracts showed inhibitory effect over growth of *Streptococcus pyogenes*, *Bacillus cereus*, *Escherichia coli*, *Acinetobacter baumannii*, *Neisseria gonorrhoeae* in well diffusion method. Overall, dichloromethane showed more activity against Gram negative bacteria on well diffusion. Acetone extracts showed inhibitory effect on *Streptococcus pyogenes*, *Bacillus cereus*, *Escherichia coli*, *Acinetobacter baumannii* and *Neisseria gonorrhoeae*. As in disc diffusion, it showed inhibitory effect on *Bacillus cereus*, *Escherichia coli*, *Neisseria gonorrhoeae* except on *A. baumannii*. Overall, it showed more antimicrobial activity in well diffusion method and Gram-negative bacteria. Ethanolic extracts showed inhibitory effect on *S. pyogenes* and *E. coli* in well diffusion. In disc diffusion, it showed inhibitory effect against *B. cereus*, *E. coli*, *A. baumannii*, and *N. gonorrhoeae*. To summarize, ethanolic extracts showed more antimicrobial activity in disc diffusion method and Gram-negative bacteria. Methanolic extracts showed inhibitory effect against *B. cereus* and *N. gonorrhoeae* in well diffusion method. Whereas in disc diffusion method, it showed inhibitory effect in all studied bacteria. To summarize, methanolic extracts showed more antimicrobial activity in disc diffusion method and Gram-negative bacteria. Based on the present antimicrobial study conducted, the study showed that dichloromethane extract has the highest antimicrobial activity followed by ethanolic extract, acetone extract and lastly methanolic extract. For three microorganisms (*S. pyogenes*, *A. baumannii*, and *N. gonorrhoeae*), so far, no study has been done, it is reported for the first time in our study. Extracts used in present antimicrobial study have shown overall good antimicrobial activity against these three bacteria in both disc and well diffusion.

#### 5. Conclusion

Based on antibacterial potential of different solvent extracts of *Cymbopogon citratus* stems, it is hereby concluded that dichloromethane extract of *Cymbopogon citratus* stems exerts highest inhibitory action against growth of bacterial strains of *Staphylococcus aureus*, *Streptococcus pyogenes*, *Bacillus cereus*, *Escherichia coli*, *Acinetobacter baumannii* and *Neisseria gonorrhoeae*. The high antimicrobial potential of dichloromethane extract of *Cymbopogon citratus* stems may be due to presence of flavonoidal constituents. Present study concludes high antibacterial potential of *Cymbopogon citratus* plant. To obtain deeper insight into the mechanism of action of extracts of stems of *Cymbopogon citratus* plant, further investigation on isolates of extracts should be done.

#### 6. References

1. Pahwa, S; Fuloria, N. K., Kumar, N., Singh V., Fuloria, S., 2007. Diversified beauty of *Saccharomyces boulardii*. *Pharmaceutical reviews*, 5(6).
2. Fuloria, N. K., Fuloria, S., Chia, K. Y., Karupiah, S., Sathasivam, K. (2019). Response of green synthesized drug blended silver nanoparticles against periodontal disease triggering pathogenic microbiota. *Journal of Applied Biology and Biotechnology*, 7, 46-56.

3. Fuloria, S., Fuloria, N. K., Karupiah, S., Sathasivam, K., Singh, S., Gupta, K., Jain, A., Sridevi, U., Himaja, M., Shanker, S. (2017). Synthesis & discerning of antibiotic potential of PCMX based novel azetidinones. *Acta Polonicae Pharmaceutica*, 76, 6, 171-1715.
4. Fair, R.J., and Tor, Y., 2014. Antibiotics and bacterial resistance in the 21st century. *Perspectives in Medicinal Chemistry*, 6, 25-64.
5. Subramaniyan, V., Fuloria, S., Chakravarthi, S., Aaleem, A. A. K., Jafarullah, S. M., Sekar, M., Sathasivam, K., Meenakshi, D. U., Kumari, U., Kumar, V., Seng, W. Y., Fuloria N. K., 2021. Dental Infections and Antimicrobials. *Journal of Drug and Alcohol Research*, 10(4), 1-5.
6. Ali, B., Al-Wabel, N. A., Shams, S., Ahamad, A., Khan, S. A., & Anwar, F. (2015). Essential oils used in aromatherapy: A systemic review. *Asian Pacific Journal of Tropical Biomedicine*, 5(8), 601-611.
7. Gauniya, A., Fuloria, S., Tripathi, P., Fuloria, N., Pahwa, S., Basu, S. P. (2008). Role of aromatic plants in national economy. *Pharmaceutical reviews*, 6(1).
8. Shivkanya, J., Shilpa, P., Sangita, K., Neeraj, F., 2009. Pharmacognostical studies and antibacterial activity of the leaves of *Murrayakoenigii*. *Pharmacognosy Journal*, 1(3), 211-214.
9. Sharma, P. K., Fuloria, S., Alam, S., Sri, M. V., Singh, A., Sharma, V. K., Kumar, N., Subramaniyan, V., Fuloria, N. K. (2021). Chemical composition and antimicrobial activity of oleoresin of *Capsicum annum* fruits. *Mindanao Journal of Science and Technology*, 19(1), 29-43.
10. Velu, V., Banerjee, S., Rajendran, V., Gupta, G., Chellappan, D. K., Kumar, N., Fuloria, S., Mehta, M., Dua, K., Malipeddi, H. (2021). Identification of phytoconstituents of *Tragiainvolucrata* leaf extracts and evaluate their correlation with anti-inflammatory & antioxidant properties. *Anti-inflammatory & Anti-allergy Agents in Medicinal Chemistry*, 20, 1.
11. Fuloria, S., Ru, C. S., Paliwal, N., Karupiah, S., Sathasivam, K., Singh, S., Gupta, K., Fuloria, N. K. (2020). Response of biogenic zinc oxide nanoparticles against periimplantitis triggering non periodontal pathogen. *International Journal of Research in Pharmaceutical Sciences*, 11, 3, 3889-3896.
12. Hema, D., Shankar, J., Ishwin, S., Vetriselvan, S., Gayathiri, S., Shereenjeet, Kaur., & Yaashini, A. (2012). Wound healing activity of terminalia arjunain albino wistar rats. *International Journal of Phytopharmacology*, 3(3), 234-240.
13. Kaur, S., Vetriselvan, S., Hemah, C., Gayathiri, S., Yaashini, A., Singh, I., & Shankar, J. (2012). Hepatoprotective activity of aqueous extract of *Picrorhizakurroa* in carbon tetrachloride (ccl4) induced hepatotoxicity in albino wistar rats. *Int J Pharm Ther*, 3, 207-214.
14. Yaashini, A., Shankar, J., Ishwin, S., Shereenjeet, K., Hema, D., & Vetriselvan, S. (2012). Hepatoprotective activity of *Bacopamonnieri* extract in Ethanol induced Hepatotoxicity in Albino Rats. *International Journal of Pharmacy and Therapeutics*, 3 (3), 259-266.
15. Jothi, S., Vetriselvan, S., Gayathiri, S., Ishwin, S., Shereenjeet, G., Devi, C. H., & Yaashini, A. (2012). Comparative evaluation of antiinflammatory activity of extract of *Curcuma longa* and standard drug in carrageenan induced paw edema model using albino Wister rats. *Internat J BiologPharmaceut Res*, 3, 538-544.
16. Singh, I., Vetriselvan, S., Shankar, J., Gayathiri, S., Hemah, C., Shereenjeet, G., & Yaashini, A. (2012). Hepatoprotective activity of aqueous extract of *Curcuma longa* in ethanol induced hepatotoxicity in albino Wistar rats. *Int J Phytopharmacol*, 3(3), 226-233.
17. Subramaniyan, V., Saminathan Kayarohanam, A. K. J., & Kumarasamy, V. (2019). Impact of herbal drugs and its clinical application. *International Journal of Research in Pharmaceutical Sciences*, 10(2), 1340-1345.
18. Hang, C. Z., Fuloria, N. K., Hong, O. J., Kim, C. B., Ting, B. Y. S., Ru, C. S., Fuloria, S. (2020). Biosynthesis of DLLAE blended silver nanoparticles and their response against periodontitis triggering bacteria. *International Journal of Research in Pharmaceutical Sciences*, 11, 1849-1856.
19. Fuloria, S., Subramaniam G., Ying, T. S., Vijpuri N., Fuloria N. K., Sikarwar S. M., Kaveti B., Sundram K., Vijayabalan S. Discerning the antioxidant and microkinetic potential of eucalyptus viminalis labill extracts. *Rapports De Pharmacie*, 2 (4), 304.
20. Vetriselvan, S., Shankar, J., Gayathiri, S., Ishwin, S., Devi, C. H., Yaashini, A., & Sheerenjet, G. (2012). Comparative evaluation of in vitro antibacterial and antioxidant activity using standard drug and polyherbal formulation. *Int J Phytopharm*, 3, 112-6.
21. Vetriselvan, S., Shankar, J., Gayathiri, S., Ishwin, S., Devi, C. H., Yaashini, A., & Sheerenjet, G. (2012). Comparative evaluation of in vitro antibacterial and antioxidant activity using standard drug and polyherbal formulation. *Int J Phytopharm*, 3, 112-6.
22. Subasini, U., Thenmozhi, S., Sathyamurthy, D., Vetriselvan, S., Victor Rajamanickam, G., & Dubey, G. P. (2013). Pharmacognostic and phytochemical investigations of *Dioscorea bulbifera* L. *International Journal of Pharmacy & Life Sciences*, 4(5).
23. Srivastava, V., Dubey, S., and Mishra, A. A., 2013. Review on lemon grass: agricultural and medicinal aspect. *International Research Journal of Pharmacy*, 4(8), 42-4.

24. Mirghani, M.E., Liyana, Y., and Parveen, J., 2012. Bioactivity analysis of lemongrass (*Cymbopogon citratus*) essential oil. *International Food Research Journal*, 19(2), 569.
25. Tajidin, N.E., Ahmad, S.H., Rosenani, A.B., Azimah, H., and Munirah, M., 2012. Chemical composition and citral content in lemongrass (*Cymbopogon citratus*) essential oil at three maturity stages. *African Journal of Biotechnology*, 11(11), 2685-93.
26. Fuloria, N. K., Fuloria, S., Sathasivam, K., Karupiah, S., Balaji, K., Jin, L.W., Jade, O. D., Jing, I. C. J. (2017). Synthesis and discerning of antimicrobial potential of novel oxadiazole derivatives of chloroxylenol moiety. *Acta Poloniae Pharmaceutica*, 74(4), 1125.
27. Fuloria, S., Fuloria, N. K., Balaji, K., Karupiah, S., Sathasivam, K., Jain, A., Sridevi, U., Himaja, M. (2016). Synthesis and discerning of the antimicrobial potential of new azomethines derived from chloroxylenol. *Indian Journal of Heterocyclic Chemistry*, 26 (1-2), 95-99.
28. Fuloria, S., Fuloria, N. K., Balaji, K., Karupiah, S., Sathasivam, K., Jain, A., Sridevi, U., Himaja, M. (2016). Synthesis, characterization and antimicrobial evaluation of 2-phenylpropanoic acid derived new oxadiazoles. *Indian Journal of Heterocyclic Chemistry*, 26 (01-02), 37-42, 2016.
29. Fuloria, S., Fuloria, N. K., Balaji, K., Karupiah, S., Sathasivam, K., Jain, A., Sridevi, U., Himaja, M., Shanker S. (2016). Synthesis and evaluation of antimicrobial potential of novel oxadiazoles derived from tolyloxy moiety. *Indian Journal of Heterocyclic Chemistry*, 26 (03-04), 101-105.
30. Fuloria, S., Fuloria, N. K., Balaji, K., Sundram, K. M. (2015). Evidences of Antitubercular Potential of Novel Thiazolidinone Derivatives Bearing Chloroxylenol Moiety. *Malaysian Journal of Pharmacy*, 2 (1), 67.
31. Redfern, J., Kinninmonth, M., Burdass, D., and Verran, J., 2014. Using soxhlet ethanol extraction to produce and test plant material (essential oils) for their antimicrobial properties. *Journal of Microbiology & Biology Education*, 15(1), 45.
32. Calabrese, L. (2005). The use and methods of making a herbarium/plant specimens. A Herb Society of America Guide. *The Herb Society of America*, 6.
33. Bajaj, S., Fuloria, S., Subramaniyan, V., Meenakshi, D. U., Wakode, S., Kaur, A., Bansal, H., Manchanda, S., Kumar, S., Fuloria, N. K. (2021). Chemical characterization and anti-inflammatory activity of phytoconstituents from *swertia alata*. *Plants*, 10(6), 1109.
34. Sharma, P. K., Fuloria, S., Ali, M., Singh, A., Kushwaha, S. P., Sharma, V. K., Subramanyan, V., Fuloria, N. K. (2021). Isolation of new phytometabolites from *Alpinia galanga* Wild rhizomes. *Pakistan Journal of Pharmaceutical Sciences*, 34, 4, 1397-1401.
35. Fuloria, N. K., Fuloria, S., Sharma, V. K., Ali, M., Singh, A., & Sharma, P. K. (2020). Isolation of new diterpene from methanolic extract of *Capsicum annuum* Linn. fruits. *Pharmacognosy Magazine*, 16(72), 730.
36. Fuloria, S., Wei, L. T., Karupiah, S., Subramaniyan, V., Gellknight, C., Wu, Y. S., Kayarohanam, S., Fuloria, N. K. (2020). Development and Validation of UV-visible method to determine gallic acid in hydroalcoholic extract of *Erythrina fusca* leaves. *International Journal of Research in Pharmaceutical Sciences*, 11(4), 6319-6326.
37. Vakiloddin, S., Fuloria, N., Fuloria, S., Dhanaraj, S. A., Balaji, K., & Karupiah, S. (2015). Evidences of hepatoprotective and antioxidant effect of *Citrullus colocynthis* fruits in paracetamol induced hepatotoxicity. *Pakistan journal of pharmaceutical sciences*, 28(3).
38. K. Balaji, L. H. Ni, B. Rajindran, M. S. Sikarwar, N. K. Fuloria, and S. Fuloria. Determination of Total Phenolic, Flavonoid Content and Antioxidant Activity of *Terminalia Chebula* (Fruit). *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 6(2), 413.
39. Jaju, S., Indurwade, N. H., Sakarkar, D. M., Ali, M., Fuloria, N. (2011). Antidiabetic & anti-inflammatory studies of *Alpinia galanga* rhizome. *Asian Journal of Chemistry*, 23, 3.
40. Jaju, S., Indurwade, N. H., Sakarkar, D. M., Ali, M., Fuloria, N. (2010). Isolation of  $\beta$ -sitosterol diglucosyl caprate from *Alpinia galanga*. *Pharmacognosy Research*, 2(4), 264.
41. Jaju S. B., Fuloria, N. K., Bhargav, K. L. (2010). Anthelmintic activity of *Ferula foetida* Regel. *Antiseptic*, 107(2), 97.
42. Gayathiri, S., Vetrivelan, S., Shankar Jothi, I. S., Hemah Devi, S. K., & Yaashini, A. (2012). Hepatoprotective activity of aqueous extract of *Hippophae rhamnoides* L. in carbon tetrachloride induced hepatotoxicity in albino wistar rats. *International journal of biological & pharmaceutical research*, 3(4), 531-537.
43. Vetrivelan, S., Rusliza, B., Subasini, U., & Velmurugan, C. (2012). Wound Healing activity of ethanolic polyherbal extract in wistar rats. *International Journal of Research in Pharmaceutical and Nano Sciences*, 1(1), 19-26.
44. Vetrivelan, S., Victor, R., Parimala, D., & Arun, G. (2010). Comparative evaluation of hepatoprotective activity of *andrographis paniculata* and *Silymarin* in ethanol induced hepatotoxicity in albino wistar rats. *Der Pharmacia Lettre*, 2(6), 52-59.



45. Vetrivelan, S., Subasini, U., Velmurugan, C., Muthuramu, T., & Revathy, J. (2013). Anti-inflammatory activity of *Cucumis sativus* seed in carrageenan and xylene induced edema model using albino wistar rats. *Int J Biopharm*, 4(1), 34-37.
46. Vetrivelan, S., Felix, A., Magendran, R., Ponnaiyakannan, S., Prabakaran, T., Jothi, S., & Davan, R. (2012). The phytochemical screening and the anti-ulcer activity of methanolic extract of *Ixora coccinea* Linn leaf. *J Pharm Res*, 5(6), 3074-3077.
47. Subramaniyan, V., Shaik, S., Bag, A., Manavalan, G., & Chandiran, S. (2018). Potential action of *Rumex vesicarius* (L.) against potassium dichromate and gentamicin induced nephrotoxicity in experimental rats. *Pakistan journal of pharmaceutical sciences*, 31(2), 509-516.
48. Vetrivelan, S., & Velmurugan, P. (2017). Anti-arthritis activity of aqueous extract of *Achyranthes aspera*. *Asian Journal of Pharmaceutical and Clinical Research*, 10(10), 372-375.
49. Venkateshan, S., Subramaniyan, V., Chinnasamy, V., & Chandiran, S. (2016). Anti-oxidant and anti-hyperlipidemic activity of *Hemidesmus indicus* in rats fed with high-fat diet. *Avicenna journal of phytomedicine*, 6(5), 516.
50. Subramaniyan, V., & Middha, A. (2016). Chronic ethanol consumption-induced hepatotoxicity and protective effect of *Boswellia serrata*. *National Journal of Physiology, Pharmacy and Pharmacology*, 6(2), 170-174.
51. Chinnasamy, V., Subramaniyan, V., Chandiran, S., Kayarohanam, S., Kannian, D. C., Velaga, V. S. S. R., & Muhammad, S. (2019). Antiarthritic Activity of *Achyranthes Aspera* on Formaldehyde-Induced Arthritis in Rats. *Open access Macedonian journal of medical sciences*, 7(17), 2709.
52. Jaju, S. B., Indurwade, N. H., Sakarkar, D. M., Fuloria, N. K., Ali, M. D., Das, S., & Basu, S. P. (2009). Galangoflavonoid isolated from rhizome of *Alpinia galanga* (L) Sw (Zingiberaceae). *Tropical Journal of Pharmaceutical Research*, 8(6), 545-550.
53. Jaju, S., Indurwade, N., Sakarkar, D., Fuloria, N., & Ali, M. (2009). Isolation of galangogalloside from rhizomes of *Alpinia galanga*. *International Journal of Green Pharmacy (IJGP)*, 3(2), 144-147.
54. Jaju, S. B., Indurwade, N. H., Sakarkar, D. M., Fuloria, N. K., Ali, M. (2009). Isolation of  $\beta$ -sitosterodiglucoside and  $\beta$ -sitosteryl arabinoside from rhizomes *Alpinia galanga*. *Asian Journal of Chemistry*, 21(3), 2350-2356.
55. Jaju, S. B., Indurwade, N. H., Sakarkar, D. M., Fuloria, N. K., Ali, M. (2009). Galangoisoflavonoid isolated from rhizomes of *Alpinia galanga*. *Pharmacognosy Magazine*, 5(19), 209-12.
56. Jaju, S. B., Indurwade, N. H., Sakarkar, D. M., Fuloria, N. K., Ali, M. (2009). Linolein-2-Stearin Phosphate and Linolenic Acid  $\beta$ -D-Glucoside: The Newer Isolates of *Alpinia galanga* Rhizomes. *Asian Journal of Chemistry*, 21(5), 3892-3896.
57. Jaju, S. B., Indurwade, N. H., Sakarkar, D. M., Fuloria, N. K., Ali, M. (2009). Linoleic acid isolated from *Alpinia galanga*. *Nigerian Journal of Natural Products and Medicine*, 12, 310-31.
58. Fuloria, S., Fuloria, N. K., Gupta, R. (2014). Synthesis and Antimicrobial Profile of Newer Schiff Bases and Thiazolidinone Derivatives. *International Journal of Chemical, Nuclear, Metallurgical and Materials Engineering*, 8(12).
59. Gupta, R., Fuloria, N. K., Fuloria, S. (2012). Synthesis & antimicrobial profile of some new heterocycles bearing thiazole moiety. *Southern Brazilian Journal of Chemistry*, 20(20), 61.
60. Husain, A., Varshney, M., Percha, V., Fuloria, N. (2013). Synthesis, characterization and biological evaluations of some 5-(substituted amino alkyl)-2-[(1, 3- benzothiazole-2-yl)]-thiazolidine-4 one Mannich bases as potent antibacterial agent. *Journal of Applied Pharmaceutical Science*, 3(4), 135-138.
61. Fuloria, S., Fuloria, N. K., Yi, C. J., Khei, T. M., Joe, T. A., Wei, L. T., Karupiah, S., Paliwal, N., Sathasivam, K. (2019). Green Synthesis of Silver Nanoparticles Blended with Citrus Hystrix Fruit Juice Extract and their Response to Periodontitis Triggering Microbiota. *Bulletin of Environment, Pharmacology and Life Sciences*, 8(7), 112-123.
62. Chauhan, V., Fuloria, N. K., Fuloria, S. (2012). Synthesis, characterization and comparative screening of some newer 2-phenyl indole and 5-chloro-2-phenyl indole derivatives. *Southern Brazilian Journal of Chemistry*, 20 (20), 69-76.
63. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2008). Synthesis & antimicrobial studies of novel imines and oxadiazoles. *Southern Brazilian Journal of Chemistry*, 16(16), 11-22.
64. Fuloria, S., Fuloria, N. K., Hong, O. J., Kim, C. B., Ting, B. Y. S., Karupiah, S., Paliwal, N., Kumari, U., Sathasivam, K. (2020) K. Synthesis of SNPs of corn silk agrowaste and their bioactivities. *Asian Journal of Chemistry*, 32, 6, 1497-1504.
65. Jaju, S., Pahwa, S., Fuloria, N. (2009). Phytochemical and antimicrobial activity of stem and leaves of *Desmodium gangeticum* Linn. *Hamdard medicus*, 52(4), 131-135.
66. Chigurupati, S., Fuloria, N. K., Fuloria, S., Karupiah, S., Veerasamy, R., Nemala, A. R., Yi, L. J., Ilan, A. X., Shah, S. A. A. (2016). Synthesis and antibacterial profile of novel azomethine derivatives of  $\beta$ -phenylacrolein moiety. *Tropical Journal of Pharmaceutical Research*, 15(4), 821-826.

67. Balouri, M., Sadiki, M., and Ibsouda, S.K. (2016). Methods for in vitro evaluating antimicrobial activity: A review. *Journal of Pharmaceutical Analysis*, 6(2), 71-9.
68. Devillers, J., Steiman, R., and Seigle-Murandi, F. (1989). The usefulness of the agar-well diffusion method for assessing chemical toxicity to bacteria and fungi. *Chemosphere*, 19(10-11), 1693-700.
69. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2008). Synthesis, characterization and biological studies of novel imines and azetidinone derivatives of haloaryloxy moiety. *Asian Journal of Chemistry*, 20(6), 4891-4900.
70. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2009). Synthesis, characterization and antimicrobial evaluation of novel imines and thiazolidinones. *ActaPoloniaePharmaceutica-Drug Research*, 66(2), 141-146.
71. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2009). Antimicrobial evaluation of imines and thiazolidinones derived from 3-phenyl propanehydrazide. *ActaPoloniaePharmaceutica-Drug Research*, 66(4), 371-377.
72. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2008). Synthesis, characterization and biological studies of neweschiff bases and azetidinones derived from propionic acid derivatives. *Asian Journal of Chemistry*, 20(8), 6457-6462.
73. Gupta, R., Fuloria, N. K., Fuloria, S. (2013). Synthesis and antimicrobial profile of some newer 2-amino-thiazole derivatives. *Turkish Journal of Pharmaceutical Sciences*, 10(3).
74. Chinni, S. V., Gopinath, S. C. B., Anbu, P., Fuloria, N., Fuloria, S., Mariappan, P., Krusnamurthy, K., Reddy, L. V., Ramachawolran, G., Sreeramanan, S., Sumitha, S. (2021). Characterization and antibacterial response of silver nanoparti-cles biosynthesized using *Coccinia indica* leaves ethanolic extract. *Crystals*, 11(2), 97.
75. Tepe, B., Daferera, D., Sokmen, A., Sokmen, M., and Polissiou, M. (2005). Antimicrobial and antioxidant activities of the essential oil and various extracts of *Salvia tomentosa* Miller (Lamiaceae). *Food chemistry*, 90(3), 333-40.
76. Gupta, R., Fuloria, N. K., Fuloria, S. (2013). Synthesis and antimicrobial activity evaluation of some schiff's bases derived from 2-aminothiazole derivatives. *Indonesian Journal of Pharmacy*, 24 (1), 35-9.
77. Fuloria, N. K., Singh, V., Shaharyar, M., Ali, M. (2009). Synthesis and antimicrobial evaluation of newer oxadiazoles derived from phenylpropionohydrazides. *Molecules*, 14(5), 1898-1903.
78. Fuloria, S., Ying, C.Y., Xuan, K. Y., Jun, C. T., Karupiah, S., Kumari, U., Fuloria, N. K., (2020). Determination of total phenolic content and antimicrobial potential of different extracts of *Citrus hystrix* DC leaves. *Bulletin of Environment, Pharmacology and Life Sciences*, 9(8), 112-116.