## Ameliorative Potentials Of Ethanolic Leaves Extracts Of *Cassia occidentalis* And *Pithecellobium dulce* On Diabetes Associated Metabolic Alterations In Streptozotocin-Induced Diabetic Rats: A Comparative Study

Badri. Rajarajeswari<sup>1</sup>; Bhogavalli. Praveen Kumar<sup>2</sup>; Audipudi. Amurtha valli<sup>3</sup>\*

<sup>1</sup>Lecturer, Department of Botany, DKW College, Nellore-524003, Andhra Pradesh, India <sup>2</sup>Biology Faculty, Department of biology, Master JEE IIT academy, Kalpakkam, Tamil Nadu, India <sup>3</sup>Associate Professor, Department of Microbiology, Acharya Nagarjuna University, Guntur, Andhra Pradesh, India

Corresponding Author's Email ID: audipudiamrita@gmail.com

### Abstract:

A comparative analysis was performed in this study to detect the ameliorative potentials of Ethanolic leaves extracts (EL) of *Cassia occidentalis* and *Pithecellobium dulce* medicinal plants by oral administration of 200mg/kg bw and 400mg/kg bw individually and in combination on streptozotocininduced diabetic rats. In diabetic rats, the treatment of EL extracts of both plant species (1:1) in combination of 400 mg/kg bw doses and time following significantly reduced blood glucose level  $(101.67\pm 9.38 \text{ mg/dl})$  and body weight  $(201.67\pm 9.38 \text{ g})$  to normal. Individual plant EL extracts also demonstrated a successful response in diabetic rats at doses of 200 and 400 mg/kg bw in restoring insulin and hepatic glycogen levels to near-normal levels. In both normal and diabetic rats, total cholesterol, triglycerides, AST, ALT, carbohydrates metabolic enzymes (SOD, CAT, and GPx) were measured. In STZ-diabetic rats, oral administration of EL extracts from both plants in combination significantly and dose-dependently normalized the above serum and liver parameters (p<0.05). Histopathological analysis revealed a protective and restorative effect.

Key words: Antidiabetic activity, Streptozotocin, Insulin, Ethanolic Extracts, Pancreas.

### 1] Introduction:

Diabetes mellitus (DM) is a highly variable, adlerian endocrine disorder characterized by chronic hyperglycemia that persists due to defects in insulin secretion, action, or both <sup>[1]</sup>. DM is marked by the appearance of hypercholesterolemia, hypertriglyceridemia, and low serum high-density lipoprotein cholesterol (HDL-C) in diabetics compared to normal individuals <sup>[2-3]</sup>. Chronic hyperglycemia causes microvascular impediments that affect the nerves, kidneys, and eyes, as well as increasing the risk of cardiovascular disease <sup>[4]</sup>.

According to the International Diabetes Federation (IDF), the global burden of diabetes was 366 million in 2011 and is expected to rise to 552 million by 2030. Despite newer and more successful treatment strategies, new diagnostic devices, strict glycemic targets, better treatment guidelines, and increased disease awareness, baseline glycosylated haemoglobin remains relatively high in diabetic subjects <sup>[5]</sup>. In India, this disorder is at an alarmingly high level when compared to the majority of developed countries. By 2025, India is expected to have the most people with Diabetes Mellitus in the world <sup>[6]</sup>. Despite advances in disorder understanding and management, the disease's mortality and morbidity are increasing <sup>[7]</sup>.

Anti-diabetic medications currently available, such as sulfonylureas, biguanides,  $\beta$ - glucosidase inhibitors, thiazolidinediones, and insulin, are frequently associated with undesirable side effects or a decrease in response after prolonged use <sup>[1]</sup>. With today's diabetes drugs, achieving better glycemic control with minimal side effects and ease of access remains a challenge. Nowadays, the use of medicinal plants is increasing because treating diabetes with phytotherapy is affordable, easily accessible, less expensive, very effective, and has fewer side effects <sup>[8]</sup>. Traditional medicine is used for primary health care by approximately 70–90% of the population in developed and developing countries because it is relatively inexpensive and has fewer side effects <sup>[9]</sup>. Natural products are important in the discovery of new therapeutic agents and have received a lot of attention as sources of bioactive substances such as antioxidants, hypoglycemic and hypolipidemic agents <sup>[10]</sup>. Because herbal preparations are regarded as a primary source of modern medicine, these facts have encouraged the

expansion of the frontiers of scientific evaluation of the hypoglycemic properties of various plant species. <sup>[11-12]</sup>.

*Cassia occidentalis* and *Pithecellobium dulce* are members of the same Fabaceae family and have been shown to have a variety of therapeutic effects <sup>[13-14]</sup>. Taking traditional claims and reported activities into account, *C. occidentalis* and *P. dulce* have been studied for their Diabetes Associated Metabolic Alterations in Streptozotocin –Induced Diabetic Rats. In light of this, the current study was undertaken to determine the synergistic anti-diabetic activity of the combination of these plants and compare its potential in streptozotocin induced diabetic rats <sup>[15]</sup>.

### 2] Materials and Methods:

**Plants collection and Authentication:** Dr. A. Amurtha Valli, Associate Professor, Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur, Andhra Pradesh, India, authenticated *C. occidentalis* and *P. dulce* plant materials collected from local village areas of Nellore district, Andhra Pradesh state. Before being shade dried for 10 days, the plant leaves were thoroughly washed, first with tap water and then with distilled water, to remove any debris or dust particles. The dried leaves were ground to a fine powder in a sterile mixer grinder for 5 minutes before being stored at room temperature in airtight plastic containers.

**Preparation of the Leaf Extract:** 500 g of previously prepared powder was mixed with 1500 ml of the solvent ethanol and left undisturbed overnight at 10°C. The solution was then allowed to stand in a boiling water bath for 1-2 hours, with occasional shaking, before being left undisturbed for 24 hours. The extract was concentrated using a rotaevaporator under vacuum below 40°C after being filtered through sterile Whatman No.1 filter paper <sup>[16-17]</sup>. The dried extract was then exposed to UV light for 24 hours before being tested for sterility on nutrient agar plates and stored in labeled sterile bottles in a freezer at 4°C until further use <sup>[18]</sup>.

**Preliminary Phytochemical Screening:** To analyze the secondary metabolites (phytochemical) constituents present in the various solvent extracts, various standard protocols were used <sup>[19-21]</sup>.

Animal Collection: Adult healthy albino Wistar rats (175 to 250 g) of both sexes were obtained from the Animal House, Ratnam Institute of Pharmacy, Nellore, Andhra Pradesh, India, and housed in the Department of Pharmacology under standard husbandry conditions ( $30^{\circ}C \pm 5$ ; 60-70% relative humidity and 12 hrs day night cycle). They were fed a standard rat pellet diet and were given unlimited access to water. All animal procedures were approved by the Institutional Animal Ethical Committee, and the experiment was carried out in accordance with the CPCSEA Guidelines.

Acute Oral Toxicity Study of the EL Extracts: An acute oral toxicity study of the EL extracts of *C. occidentalis* and *P. dulce* was carried out in accordance with the guidelines of the Organization for Economic Cooperation and Development (OECD) <sup>[22]</sup>. Prior to oral dosing, the rats were fasted overnight and given only water. The extract was then administered orally at various dose levels of 100, 200, 500, 1000, 1500, and 2000 mg/kg of body weight. The rats were monitored continuously for 24 hours for behavioral and adverse changes, and then for lethality.

**Diabetes induction and experimental design:** The experimental rats were fasted overnight, and Streptozotocin (STZ) was weighed individually for each rat before being injected intraperitoneally with STZ (50 mg/kg bw) in freshly prepared 0.1M of cold citrate buffer (pH 4.5) <sup>[23-24]</sup>. The rats were given free access to food and water after 30 minutes of injection. After 48 hours of STZ administration, the blood glucose level was estimated. Animals with fasting blood glucose levels greater than 200-350 gm/dl were considered diabetic and were chosen for the next activity. 48 hours after STZ injection, treatment with EL extracts *C. occidentalis* and *P. dulce* was initiated. The chosen animals were divided into nine groups of six each.

**Group I:** A normal control group that did not receive STZ injections received a daily dose of 0.3 ml of 2.5 percent Tween 80 (vehicle) p.o.

Group II: Diabetic control with a single intravenous dose of 50 mg/kg bw STZ.

**Group III:** Diabetic control with a daily dose of 5 mg/kg bw Glibenclamide in 2.5 percent Tween 80 p.o.

Group IV: Diabetic control with a daily dose of 200 mg/kg bw ELCO in 2.5 percent Tween 80 p.o.

Group V: Diabetic control with 400 mg/kg bw ELCO in 2.5 percent Tween 80 p.o. daily dose.

Group VI: Diabetic control with a daily dose of 200 mg/kg bw ELPD in a 2.5 percent Tween 80 p.o.

solution.

**Group VII:** Diabetic control with 400 mg/kg bw ELPD daily dose in 2.5 percent Tween 80 p.o. **Group VIII:** Diabetic control with a daily dose of 200 mg/kg bw ELCO & ELPD (1:1) in 2.5 percent Tween 80 p.o.

Group IX: Diabetic control with 400 mg/kg bw ELCO & ELPD (1:1) in 2.5 percent Tween 80 p.o.

**Biochemical Assays:** A commercially available glucose oxidase kit was used to analyze fasting blood glucose levels (Span Diagnostics Ltd, Surat, India). The Elisa kit from Stat Diagnostics, Mumbai, was used to measure plasma insulin levels. The liver was immediately dissected, washed in ice-cold saline to remove blood, and homogenized in 0.1M Tris –HCl buffer, pH 7.4. The supernatant was tested for carbohydrate metabolic enzyme activity using Trinder (1969) <sup>[25]</sup> (Hexokinase), King (1965) <sup>[26]</sup> (Glucose-6-Phosphatase), and Gancedo and Gancedo (1971) <sup>[27]</sup> (Fructose-1,6 Bisphosphates). Morales *et al.* (1975) <sup>[28]</sup> Total cholesterol (Parekh and Jung, 1970) <sup>[29]</sup>, and Total Triglycerides (Foster and Dunn, 1973) <sup>[29]</sup>. Enzymes from the liver (AST, ALT) (Reitmann and Frankel, 1957) <sup>[30]</sup>. Antioxidant enzymes (SOD) (Kakkar *et al.*, 1984) <sup>[31]</sup>, CAT (Sinha, 1972) <sup>[32]</sup>, Rotruck *et al.*, 1973) <sup>[33]</sup> (GPx).

**Histopathological studies:** At the end of the experiments, the animals are sacrificed under sedation (Isoflurane). The liver and pancreas of one animal from each group were then excised, washed with normal saline, and stored in 10% formalin solution for histopathological analysis. The tissue was washed, dehydrated in alcohol, cleared in xylene, and paraffin blocks were created. A semi-automated rotary microtome was used to cut serial sections of 5 m thickness. The sections were deparaffinized with xylene and rehydrated in descending grades of ethanol before being stained with haematoxylin and eosin and examined under a microscope.

**Statistical analysis:** The results are expressed as the mean SEM of the number of experiments. To determine the significance of changes, the data was subjected to analysis of variance (one way ANOVA), followed by Dunnett's test.

### 3] Results and Discussion:

As a first step, a preliminary phytochemical analysis was performed on both EL leaves extracts of *C. occidentalis* and *P. dulce* medicinal plants, both of which are members of the Fabaceae family. The presence of alkaloids, flavonoids, anthraquinones, tannins, proteins, and terpinoids was discovered **(Table 1)**. Acute oral toxicity studies on both plant species revealed no behavioral changes in the first 24 hours at doses less than 2000mg/kg bw. Normal rats treated with ET extract in the range of 100-1500 mg/kg bw for both plants had a 100% survival rate.

Diabetes is a metabolic disorder that affects a large proportion of the world's population. Depending on the type and severity of diabetes, treatment is based on the variable use and combination of diet, antidiabetic oral agents (metformin, sulphonylureas, acarbose, and thiazolidinediones), and insulin or its analogues. Traditional diabetes treatments have many flaws, such as unfavorable side effects and a high rate of secondary complications. In general, there is little scientific knowledge on the specific mechanisms of action in the treatment of diabetes, but most plants have been found to contain active ingredients such as flavonoids, alkaloids, glycosides, terpenoids, and so on, which have anti-diabetic properties <sup>[35, 36]</sup>. The presence of flavonoids, alkaloids, glycosides, polyphenols, tannins, saponins, phytosterols, and triterpenes in the EL leaves extract was revealed by phytochemical analysis (**Table 1**). Diabetes prevention and treatment rely heavily on phytocompound-based strategies <sup>[37]</sup>. Polyphenolic compounds, such as flavonoids, help to increase plasma antioxidant capacity, lower oxidative stress markers, and lower total and LDL cholesterol <sup>[38]</sup>.

A growing body of evidence suggests that different dietary phytophenols may influence carbohydrate metabolism at multiple levels <sup>[39]</sup>. Phytochemicals are compounds with various biological properties that enable plants to cope with environmental challenges such as radiation and toxins <sup>[40]</sup>. They are bioactive compounds (secondary metabolites) found in plants that work in conjunction with nutrients and dietary fibres to provide disease protection <sup>[41]</sup>. The presence of biologically active ingredients in individual plant EL extracts and in combination doses could explain the observed pharmacological effects.

ISSN: 0975-3583, 0976-2833 VOL 12, ISSUE 03, 2021

	dulce (PD).				
S.N0	Phytochemicals	Test's Applied	ELCO	ELPD	
1	Alkaloids	a) Mayer's Test b) Dragendroff's Test c) Wagner's Test	++++	+++	
2	Saponins	Foam Test	-	-	
3	Flavanoids	a) Ferric Chloride Test b) Shinoida's Test	++++	++	
4	Anthraquinones	Borntrager's Test	++	++	
5	Tannins	a) Ferric Chloride Test b) Lead Acetate Test	++	++	
6	Sterols	a) Salkowski's Test b) Libermann-Burchards Test	-	+	
7	Sugars	a) Barfoed's Test b) Molish's Test	++	++	
8	Cardiac Glycosides	a) Borntrager's Test b) Balget's Test	-	-	
9	Lipds and Oils	Saponification Test	+	+	
10	Coumarins	Sodium Hydroxide Test	++	_	
11	Proteins	a) Biuret's Test b) Ninhydrin Test	+	+	
12	Terpinoids	Salkowski Test	++	+++	

Table 1: Comparative Phytoconstituents analysis in EL extracts of *C occidentalis* (CO) and *P. dulce* (PD).

In rats, diabetes was induced by a single intraperitoneal injection of STZ. Animals with severe hyperglycemia (glucose >200 mg/dl) were compared to rats with normal blood sugar levels after 48 hours. Group I rats had normal blood glucose levels and were classified as Normal, whereas Groups II- IX were classified as diabetic. For 21 days, ELCO 200 mg/kg bw and 400 mg/kg bw, ELPD 200 mg/kg and 400 mg/kg, and a combined plant extract (1:1) of ELCO and ELPD at doses of 200 mg/kg and 400 mg/kg bw were administered orally. **Table 2** compares the effect of EL extracts of ELCO, ELPD, and combined plant extract (1:1) of ELCO and ELPD at doses of 200 mg/kg on non- diabetic rats' mean fasting blood sugar to diabetic rats' mean fasting blood sugar. Blood glucose levels were significantly lower after treatment with EL plant extract. At a dose of 400 mg/kg, the combined plant extract of ELCO and ELPD showed significant and promising signs of anti-diabetic activity against STZ-induced diabetic rats, and the effect was comparable to that of the standard drug glibenclamide (5 mg/kg).

Table 2: Effect of STZ, Standard and EL extracts on blood glucose level in different groups of
normal and diabetic rats.

normai and diabetic rats.		1		
Grou	Pre-treatment	Post-treatment		
ps				
	Day 0	Day 7	Day 14	Day 21
	· ·	·	·	· ·
Normal control	$87.64 \pm 5.42$	$89.45\pm7.62$	$90.76\pm4.43$	$91.07\pm8.45$
Diabetic control	$289.5 \pm 6.123$	$324.5 \pm 10.47$	$380.5 \pm 14.11$	$408.5\pm15.68$
Glibenclamide 5mg/kg bw	$268.17 \pm 15.33$	$212.17 \pm 10.41$	$162.17 \pm 12.47$	$96.17\pm8.75$
200 mg/kg bw ELCO	$224.33 \pm 14.23$	$220.65\pm9.65$	$180.33 \pm 10.84$	$133.17 \pm 11.12*$
400 mg/kg bwELCO	$254.67 \pm 17.84$	$236.33 \pm 18.68$	$147.17 \pm 15.41$	$102.67 \pm 40.07 **$
200 mg/kg bwELPD	$265.33\pm9.38$	$244.67 \pm 18.12$	$194.67 \pm 14.56$	$148.83 \pm 13.74*$
400 mg/kg bwELPD	$268.45 \pm 9.36$	$235.54 \pm 9.18$	$170.23 \pm 8.38$	109.32±10.38*
200 mg/kg bw	$251.12\pm8.34$	$224.14 \pm 6.38$	$187.89\pm9.18$	$150.45 \pm 9.38$
ELCO&ELPD (1:1)				
400 mg/kg bw	$255.05\pm9.68$	$215.12 \pm 9.38$	$164.13\pm7.38$	$101.67 \pm 9.38 **$
ELCO&ELPD(1:1)				

Each value represents the mean  $\pm$  SEM, (n=6) \*P < 0.05, Vs diabetic control (ANOVA followed by Dunnett'stest). ELCO = Ethanolic leaves extract of *C.occidentalis* and ELPD = Ethanolic leaves extract of *P. dulce*.

**Fig: 1** depicts the effect of various plant extracts on the mean body weights of non-diabetic rats versus diabetic rats. Diabetes induced by STZ is associated with a characteristic loss of body weight, which is most likely due to muscle wasting and protein loss in the tissues. The extracts ELCO, ELPD, and combined plant extract (1:1) ELCO and ELPD were given orally to different groups of diabetes animals for 21 days at doses

## Journal of Cardiovascular Disease Research

### ISSN: 0975-3583, 0976-2833 VOL 12, ISSUE 03, 2021

of 200 mg/kg and 400 mg/kg. It was discovered that there is a significant increase in body weights after 21 days. The body weight of the STZ-induced untreated diabetic group decreased significantly in our study. DM was caused by STZ injection, most likely due to destruction of the  $\beta$ -cells of the pancreatic islets of Langerhans <sup>[42]</sup> Hyperglycemia in DM is caused by an excess of glucose production and a decrease in tissue utilization <sup>[13]</sup>. In comparison to diabetic control rats and rats treated with glibenclamide, oral administration of EL at a dose of 400 mg/kg for 21 days resulted in an improvement in body weight.



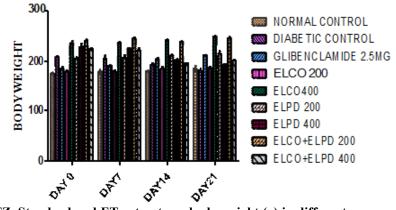


Fig 1: Effect of STZ, Standard and ET extracts on body weight (g) in different groups of normal and diabetic rats.

When plasma insulin and liver glycogen levels were studied, treated diabetic rats' levels were significantly restored to near normal levels (**Table 3**). In normal rats, plasma insulin and liver glycogen levels were  $33.06\pm 0.514$  (U/ml) and  $50.06\pm 0.215$  (mg/d wet tissue), respectively. The combined dose of 400 mg/kg bw of EL extract in (1:1) was  $34.36\pm 1.28$  (U/ml) and  $48.56\pm 1.24$  (mg/d wet tissue). Because of STZ's ability to selectively target and destroy insulin-producing pancreatic  $\beta$ - islet-cells, STZ-induced hyperglycemia is a widely used experimental model. STZ (50 mg/kg) intraperitoneal administration partially damages insulin secreting pancreatic  $\beta$ -cells by breaking the DNA strand, resulting in increased blood glucose levels and decreased endogenous insulin release. The increased plasma insulin levels in EL-treated STZ-diabetic rats could be attributed to the protection of function  $\beta$ -cells from further deterioration. Increased insulin levels may aid in the improvement of glycemic control in STZ-diabetic rats. This demonstrated the presence of active anti-diabetic active principles in both the combination and individual doses of the plant's leaf.

Groups	Parameters		
	Plasma Insulin (µU/ml)	Liver Glycogen (mg/d wet tissue)	
Normal control	$33.06 \pm 0.514$	$50.06 \pm 0.215$	
Diabetic control	$14.02 \pm 0.506$	$21.64 \pm 2.14$	
Glibenclamide (5mg/kgbw) p.o	$30.18 \pm 0.381^{*}$	45.82 ± 2.17 <sup>*</sup>	
200 mg/kgbw ELCO p.o	$25.66 \pm 0.42^*$	* 33.24 ± 3.56	
400mg/kgbw ELCO p.o	$29.50\pm0.92^*$	* 41.70 ± 1.12	
200 mg/kgbw ELPD p.o	$27.81 \pm 0.14$ *	43.11 ± 3.18 **	
400 mg/kgbw ELPD,p.o	$29.54 \pm 0.471$ *	* 47.18 ± 3.14	
200 mg/kgbw ELCO & ELPD (1:1) p.o	$30.52 \pm 0.24$ **	45.92 ± 2.22 <sup>*</sup>	
400 mg/kg ELCO & ELPD (1:1) p.o	$34.36 \pm 1.28$ *	48.56 ± 1.24 **	

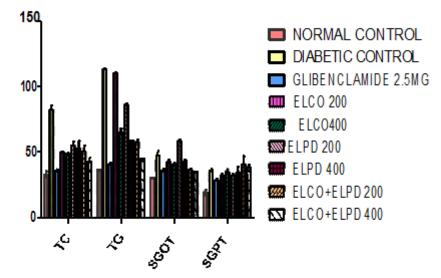
Table 3: Effect of STZ, Standard and ET extracts on plasma insulin and Liver glycogen in different groups of normal and diabetic rats.

Each value represents the mean  $\pm$  SEM, (n=6) \*P < 0.05, Vs diabetic control (ANOVA followed by Dunnett'stest). ELCO = Ethanolic leaves extract of *C. occidentalis* and ELPD = Ethanolic leaves extract of *P. dulce*.

The effect of ELCO and ELPD extracts on biochemical parameters such as total cholesterol, triglycerides, and liver enzymes SGOT and SGPT was also investigated and is depicted in Fig: 2. Diabetes is a metabolic disorder that causes changes in biochemical parameters. Triglyceride levels in 1278

diabetic rats were found to be elevated up to 113.60 mg/dl. This observation clearly demonstrated that changes in biochemical and liver function were also altered in diabetic rats. When ELCO, ELPD, and combined plant extract (1:1) of 200 mg/kg and 400 mg/kg treated rats were compared to diabetic control rats, there was a significant decrease in total cholesterol, triglyceride, SGOT, and SGPT. Hypertriglyceridemia and hypercholesterolemia are the most commonly observed lipid alterations in diabetes <sup>[43]</sup>. The presence of too many fatty acids in the bloodstream may aid in the hepatic conversion of fatty acids into phospholipids and cholesterol. These modifications are likely to result in secondary complications of diabetes, such as atherosclerosis and increased coronary heart disease. Our findings show that EL administration significantly reduced lipid profiles such as cholesterol and triglycerides.

It is important to measure the enzymes activities of aminotransferases SGOT and SGPT (AST and ALT). They were clinically and toxicologically significant because they were directly associated with the conversion of amino acids to Ketone acids are a type of acid. As a result, changes in their activities are indicative of tissue damage caused by toxicants or disease conditions. Transaminase levels in the liver are extremely high as a result of hepatocellular disorders <sup>[44]</sup>. The increase in plasma SGOT and SGPT activities suggested that diabetes may cause hepatic dysfunction due to liver necrosis <sup>[45]</sup>. When compared to the diabetic group, oral administration of EL extract and glibenclamide significantly reduces the activity of these enzymes. The return of diabetic rats' plasma AST, ALT levels to normal demonstrates that the EL extract has no negative effect on liver functions (Fig: 2).



## CHANGES IN SERUM PARAMETERS IN NORMAL AND STZ INDUCED RATS

## Fig 2: Effect of STZ, Standard and ET extracts on serum parameters in different groups of normal and diabetic rats.

Changes in the glycolytic enzyme (Hexokinase), gluconeogenesis enzyme (Glucose-6-phosphatase), and fructose 1, 6 bisphospatases were also monitored in the liver homogenate because this work is solely concerned with carbohydrate metabolism. A significant decrease in hepatic hexokinase levels was observed, with a concurrent increase in Glucose-6-phosphatase and Fructose 1, 6 bisphospatases levels (Table 4). In STZ-induced diabetic rats, the alterations were nearly normalised after treatment with a combination dose of ET (400mg/kg bw). Hepatic glucose-6-phosphatase catalyses glucose production and is essential for maintaining blood glucose homeostasis. Increased glucose-6phosphatase activity stimulates lipogenesis in diabetic rats by providing hydrogen to NADP<sup>+</sup>, forming NADPH, and enhancing fat synthesis from carbohydrates <sup>[46]</sup>, as well as increasing blood glucose levels <sup>[47]</sup>. The current study shows that hepatic glucose-6-phosphatase activity in diabetic rats was significantly higher than in normal rats, and that oral administration of EL extract and glibenclamide significantly reduced its activity and the amount of serum glucose released into the bloodstream. Fructose 1-6-phosphatase is a gluconeogenic pathway regulatory enzyme that catalyses the conversion of fructose-1, 6-bisphosphate to fructose-6-phosphate. In diabetics, activity of fructose 1-6-phosphatase increases, resulting in a decrease in glycolytic flux [48]. The oral administration of EL extract and glibenclamide reduces the activities of the gluconeogenic enzyme in diabetic rats (Table 4) by potentiating the effect of insulin release from  $\beta$ -islets of Langerhans cells, which may improve glucose utilization<sup>[49]</sup>.

Groups	Hexokinase	Glucose-6- phosphatase	Fructose-1,6-bis phosphatase
Normal control	$236.29 \pm 1.54$	1029. 8 ± 20.3	490.33 ± 18.22
Diabetic control	$138.5 \pm 2.37$	$1677.29 \pm 45.20$	$756.9 \pm 25.46^{a}$
Glibenclamide 2.5mg/kg bw	$221.34 \pm 2.21^{a}$	$1086.8 \pm 35.29^{a}$	509.3 ± 12.67
200 mg/kg bw ELCO&ELPD (1:1)	$217.67 \pm 11.13^{a}$	$1068.28 \pm 53.20^{b}$	$456.43 \pm 21.31^{b}$
400 mg/kg bw ELCO&ELPD(1:1)	$232.50 \pm 11.22^{a}$	$999.82 \pm 13.29^{\mathrm{a}}$	$473.25 \pm 1.28^{b}$

# Table 4: Effect of EL extract on carbohydrate metabolizing enzymes levels in control and STZ induced diabetic rats.

Each value is mean  $\pm$  SD for 6 rats in each group. a: p<0.05 by comparison with normal rats. b: p<0.05 by comparison with STZ diabetic rats. ELCO = Ethanolic leaves extract of *C. occidentalis* and ELPD = Ethanolic leaves extract of *P. dulce*.

The observation on antioxidant enzymes also yielded positive results, as the levels of SOD, CAT, and GPx are drastically reduced due to the effect of STZ (**Table 5**). When compared to the standard drug, diabetic rats responded well to the combination EL extracts of 200mg/kg and 400mg/kg bw. Oxidative stress is important in the development of hyperglycemia because it generates reactive oxygen species (ROS) that cause cellular injury and several deleterious effects on cellular physiology, all of which play a role in the development of secondary diabetes complications. Several studies have shown that STZ-treated  $\beta$ -cells produce oxygen free radicals and those antioxidant enzymes such as SOD and CAT are over expressed. Diabetes has been linked to decreased SOD and CAT activities in the liver. SOD is a critical defense enzyme that catalyses superoxide radical dismutation. CAT is a heme protein that catalyses hydrogen peroxide reduction and protects tissues from highly reactive hydroxyl radicals <sup>[50]</sup>. Low catalase activity has been linked to schizophrenia and atherosclerosis <sup>[51]</sup>, and it is thought that long-term oxidative stress may lead to the development of Type II diabetes. In diabetic rats, oral administration of EL extract and glibenclamide significantly increases SOD, CAT, GPx levels (**Table 5**). The EL extract's ability to restore the altered antioxidant enzymes and glutathione in STZ-induced diabetic rats suggests that it has free radical scavenging potential.

Table 5: Effect of EL extract on A	Antioxidative enzymes in	control and STZ – diabetic rats.
------------------------------------	--------------------------	----------------------------------

Groups	SOD (units/min / mg protein)	CAT (n moles/ 100 g tissue)	GPx (µg of SSH consumed/ min/mg	
			protein)	
	LIVER			
Normal control	9.17±0.77	83.41±6.72	9.52±0.69	
Diabetic control	4.12±0.29 ª	42.66±3.91 ª	4.12±0.50 ª	
Glibenclamide 5mg/kg bw	10.46±0.91 <sup>b</sup>	85.12±6.91 <sup>b</sup>	11.21±0.81 <sup>b</sup>	
200 mg/kg bw EELCO&EELPD (1:1)	9.51±0.68 <sup>b</sup>	79.44±5.12 <sup>b</sup>	9.17±0.73 <sup>b</sup>	
400 mg/kg bw EELCO&EELPD(1:1)	9.76±0.71 b	78.58±5.45 b	9.24±0.76 <sup>b</sup>	

Each value is mean  $\pm$  SD for 6 rats in each group. a: p<0.05 by comparison with normal rats. b: p<0.05 by comparison with STZ diabetic rats.

In comparison to the normal control group, histopathological examinations of liver sections in control rats revealed that the liver parenchyma was partially deformed structurally and had focal areas of necrosis with crammed blood vessels. Because the EL leaves extract combination of (Dose 200, 400 mg/kg (1:1) produced positive results in the previous study, the investigation was limited to combination treatments. The findings revealed a remarkable appearance of parenchyma with intact architecture and midzonal hepatocytes. (Fig:3). The section of control rat showed normal architecture with islet of Langerhans formed of numerous compactly arranged  $\beta$  - cells, while the section of diabetic rat showed pyknotic nuclei and dark nuclei, with few cells at the periphery having round or ovoid nuclei (Fig 4), while the section of rat treated with EL combination 200 mg/kg b.w showed shrunken nuclei and mild inflammatory cells (Fig 4d) and regeneration of  $\beta$ -cells (70 percent, compared to 1280

## Journal of Cardiovascular Disease Research

## ISSN: 0975-3583, 0976-2833 VOL 12, ISSUE 03, 2021

positive control group 30 percent), while the periphery was made up of large -cells (25 percent , compared to positive control group 65 percent ). Histopathology studies on the liver revealed that EL has a protective effect. EL treatment preserved the architecture of the liver parenchyma as well as the perivenular, periportal, and midzonal hepatocytes. The central veins and sinusoids appear unremarkably normal in comparison to the positive control group, which had partially distorted liver parenchyma. In a positive control rat, pancreas showed degeneration of islet  $\beta$ - cells and a decrease in islet cell mass. When compared to positive control rats, groups treated with EL showed regeneration of  $\beta$ -cells and an increase in islet mass. Glibenclamide (5 mg/kg) treatment of diabetic rats resulted in  $\beta$ - cell regeneration as well.

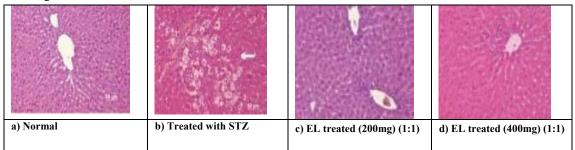
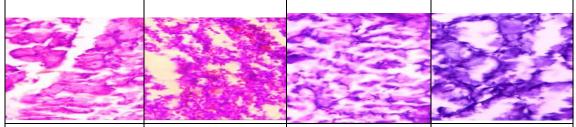


Fig 3: Histopathological panels of rat Liver prepared from 4 different groups of rats a. Control; b. Diabetic; c. Diabetic –EL combination (200 mg/kg bw); d. Diabetic – EL combination (400 mg/kg bw).



a) Normalb) Treated with STZc) EL treated (200mg)d) EL treated (400mg)Fig 4: Histopathological panels of rat pancreas prepared from 4 different groups of rats a. Control; b.Diabetic; c. Diabetic – EL combination (200 mg/kg bw); d. Diabetic –EL combination (400 mg/kg bw).

### 4] Conclusion:

In conclusion, the phytoconstituents in *C. occidentalis* and *P. dulce* exert promising anti-hyperglycemic effects in STZ-induced  $\beta$ -cell damaged diabetic rats. According to the discussion above, combined plant ethanolic extracts of *C. occidentalis* and *P. dulce* at dose (400 mg/kg) exhibited significant anti-hyperglycemic activity in STZ-induced diabetic rats. These extracts also improved parameters such as body weight and decreased serum liver parameters, suggesting that they could be useful in the treatment of diabetes. Thus, these anti-hyperglycemic agents can be investigated for the development of anti-diabetic lead molecules, and further research into mechanisms of action may result in effective diabetes treatment and control. The study suggests that the combined extract had a synergistic hypoglycemic effect, as evidenced by increased serum insulin levels and decreased serum lipid levels, and thus attribute to the combined plant extracts' therapeutic value in combating the diabetic condition in rats. However, more pharmacological and biochemical research is needed to determine the precise mechanism of hypoglycemic effects of ethanolic leaf extracts of *C. occidentalis* and *P. dulce*.

Conflict of Interest: The authors have no conflict of Interest.

Acknowledgement: The authors are very thankful to Principal and Dr. A. Amutha Valli, Associate Professor in Department of Microbiology, Acharya Nagarjuna University, for providing the lab and Materials for conducting the above research work.

#### 5] References:

[1] Pradeepa S, Subramanian S, Kaviyarasan V. Biochemical evaluation of antidiabetic properties of Pithecellobium dulce fruits studied in streptozotocin induced experimental diabetic rats. International Journal of Herbal Medicine 2013, 1 (4): 21-8

[2] Schofield JD, Liu Y, Rao-Balakrishna P, Malik RA, Soran H. Diabetes dyslipidemia. Diabetes Ther. 2016, 7: 203-19.

[3] Vijayaraghavan K. Treatment of dyslipidemia in patients with type 2 diabetes. Lipids Health Dis. 2010, 9: 144.

[4] Skyler JS, Bakris GL, Bonifacio E, Darsow T, Eckel RH, Groop L, Groop P-H, Handelsman Y, Insel RA, Mathieu C. Differentiation of diabetes by pathophysiology, natural history, and prognosis. Diabetes. 2017, 66:241-55.

[5]Vaz JA, Patnaik A. Diabetes mellitus: Exploring the challenges in the drug development process. Perspect Clin Res 2012, 3(3):109-12.

[6] Mohan V, Sandeep S, Deepa R, Shah B, Varghese C. Epidemiology of type 2 diabetes: Indian scenario diabetes. Indian J Med Res.2007, 125: 217-30.

[7] Dhanbal SP. Evaluation of therapeutic activity and development of quality control profiles for some antidiabetic herbal drugs. Indian J Pharm Educ. 2004, 8: 163-5.

[8] Tafesse TB, Hymete A, Mekonnen Y, Tadesse M. Antidiabetic activity and phytochemical screening of extracts of the leaves of Ajuga remota Benth on alloxan-induced diabetic mice. BMC Complement Altern Med. 2017, 17(1):243. doi:10.1186/s12906-017-1757-5

[9] Robinson MM, Zhang X. Traditional medicines: global situation, issues and challenges. World Med Situation. 2011, 1–14.

[10] Nabi SA, Kasetti RB, Sirasanagandla S, Tilak TK, Kumar MVJ, and Rao CA, "Antidiabetic and antihyperlipidemic activity of piper longum root aqueous extract in STZ induced diabetic rats," BMC Complementary and Alternative Medicine.2013,13(37). Doi:10.1186/1472-6882-13-37.

[11] Mihailova S, Tsvetkova A, Todorova A. Pharmacological trends in the treatment of diabetes type 2-New classes of antidiabetic drugs. Int Arch Integr Med. 2015, 2(4):223–28.

[12] Gebremeskel L, Bhoumik D, Sibhat GG, Tuem KB. In vivo wound healing and anti-inflammatory activities of leaf latex of aloe megalacantha baker (Xanthorrhoeaceae). Evid Based Complement Alternat Med. 2018, 2018:1–7. doi:10.1155/2018/5037912

[13]Laxmi Verma, P. K. Singour, P. K. Chaurasiya, H. Rajak1, R. S. Pawar, U. K. Patil. Effect of ethanolic extract of *Cassia occidentalis* Linn. for the management of alloxan-induced diabetic rats. Pharmacognosy Research. 2010, 2(3): 132-37.

14] Sukantha TA, Shubashini. K. Sripathi, Ravindran NT. Anti-Diabetic Activity Of Aqueous Extract Of Pithecellobium Dulce Benth Fruit Peel On Streptozotocin Induced Diabetic Rats. Journal of Advances in chemistry. 2016,1 2 (15): 4807-15.

15] Anu C, Abbas AM, Sohil A, Raj KS. Indian herbs result in hypoglycaemic responses in streptozotocin-induced diabetic rats. Nutr Res 2007, 27:161-8.

16] Ogundiya MO, Okunade MB, Kolapo AL. Antimicrobial activities of some Nigerian chewing sticks. Ethnobotanical Leaflets. 2006, 10: 265-71.

17] Bag A, Bhattacharyya SK, Bharati P, Pal NK, Chattopadhyay RR. Evaluation of antibacterial properties of Chebulic myrobalan (fruit of Terminalia chebula Retz.) extracts against methicillin resistant Staphylococcus aureus and trimethoprimsulphamethoxazole resistant uropathogenic Escherichia coli. African Journal of PS.2009, 3(2): 025-29.

18] Nkere CK, Iroegbu CU. Antibacterial screening of the root, seed and stem bark extracts of Picralima nitida. African Journal of Biotechnology. 2005, 4(6): 522-526.

19] Regina SN. Morphological and preliminary phytochemical studies of drug yielding herbs. Ph.D. Thesis, 2003; C.S.J.M. University, Kanpur.

20] Sofowara A. Medical Plants and Tropical Medicine in Africa. Spectrum Books LTD., Ibandan, Nigeria, 1993; pp 289.

21] Trease GE, Evan WC. Pharmacognsy. 11th edn. Brailliar Tridel Can.Macmillan Publishers.1989.

22] OECD. Organization for Economic Growth Development (OECD) guidelines for the Testing of Chemicals: Acute Oral Toxicity Up and Down Procedure (UDP); 2008.

23] Kolb H. Mouse models of insulin dependent diabetes: low-dose streptozocin- induced diabetes and nonobese diabetic (NOD) mice. Diabetes Metab Rev. 1987, 3(3):751–78. doi:10.1002/dmr.5610030308.

24] Furman BL. Streptozotocin-induced diabetic models in mice and rats. Curr Protoc Pharmacol. 2015, 70(1):5.47. 41–45.47. 20. doi:10.1002/0471141755.ph0547s70.

25] Trinder P. Determination of blood glucose using an oxidaseperoxidase system with a non-carcinogenic chromogen. J Clin Pathol. 1969, 22: 158-61.

26] King J. Practical Clinical Enzymology. D van Nostrand Co, Philadelphia, London. 1965; p. 1-301. 27] Gancedo JM, Gancedo C. Fructose-1,6-bisphosphatase, phosphofructo kinase and glucose-6phophate dehydrogenase from fermenting and non-fermenting yeast. Arch Microbiol. 1971, 76: 132-38.

28] Morales MA, Jobbagy A, Terenzi HF. Mutations affecting accumulation of glycogen in Neurospora crassa. News Lett., 1973, 20: 24-25.

29] Parekh AC, Jung DH. Cholesterol determination with ferric acetate-uranium acetate and sulfuric acid-ferrous sulfate reagents. Anal Chem. 1970, 42: 1243-47.

30] Foster LB, Dunn RT. Stable reagents for determination of serum triglycerides by colorimetric Hantzsch condensation method. Clin Chem. 1973, 18: 338-40.

31] Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxalo acetic acid, glutamic pyruvate transaminase. Am J Clin Pathol. 1957, 28: 56-63.

32] Kakkar P, Das B, and Viswanathan PN. A modified spectrophotometric assay of superoxide dismutase. Ind J Biochem Biophys. 1984, 21: 130-132.

33] Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972, 47, 389-394.

34] Rotruck JT, Pope AL, Ganther HE, et al. Selenium: biochemical role as a component of glutathione peroxidase. Science. 1973, 179: 588-590.

35] Loew D, Kaszkin M. Approaching the problem of bioequivalence of herbal medicinal products. Phytother Res. 2002, 16(8):705-11.

36] Jaiswal D, Rai PK, Watal G. Antidiabetic effect of Withania coagulans in experimental rats. Indian J Clin Biochem. 2009, 24(1):88-93.

37] Chang CL, Chen YC, Chen HM, Yang NS, Yang WC. Natural cures for type 1 diabetes: a review of phytochemicals, biological actions, and clinical potential. Curr Med Chem 2013, 20(7):899-07.

38] Hanhineva K, Törrönen R, Bondia-Pons I, Pekkinen J, Kolehmainen M, Mykkänen H, Poutanen K. Impact of dietary polyphenols on carbohydrate metabolism. Int J Mol Sci 2010, 11(4):1365-02.

39] Soory M. Nutritional antioxidants and their applications in cardiometabolic diseases. Infect Disord Drug Targets 2012, 12(5):388-401.

40] Huffman MA. Animal self-medication and ethno-medicine: exploration and exploitation of the medicinal properties of plants. Proc Nutr Soc 2003, 62(2):371-381.

41] Craig W, Beck L. Phytochemicals: Health Protective Effects. Can J Diet Pract Res 1999, 60(2): 78-84.

42] Kavalali G, Tuncel H, Goksel S, Hatemi MH. Hypoglycemic activity of Urticapilulifera in streptozotocin-diabetic rats. J Ethnopharmacol. 2002, 84: 241–5.

## ISSN: 0975-3583, 0976-2833 VOL 12, ISSUE 03, 2021

43] Ravi K, Rajasekaran S, Subramanian S. Antihyperlipidemic effect of Eugenia jambolana seed kernel on streptozotoc ininduced diabetes in rats. Food Chem Toxicol . 2005, 43:1433-9.

44] Hultcrantz R, Glaumann H, Lindberg G, Nilsson LH. Liver investigation in 149 asymptomatic patients with moderately elevated activities of serum aminotransferase. Scand J Gastroenterol. 1986, 21(1): 109-13.

45] Larcan A, Lambert H, Laprevote-Heully MC, Delorme N. Light and electron microscopic study of hepatic lesions in the course of hyperlactatemia in diabetic patients. Diabete Metab. 1979, 5(2): 103-12.

46] Bopanna KN, Kannan J, Sushma G, Balaraman R. Antidiabetic and antihyperlipidemic effects of neem seed kernel powder on alloxan diabetic rabbits. Indian J Pharmacol. 1997, 29: 162–67.

47] Venkateswaran S, Pari L. Effect of Coccinia indica on blood glucose, insulin and hepatic key enzymes in experimental diabetes. Pharmaceut. Biol. 2002, 40: 165–70.

48] Baquer NZ, Gupta D, Raju J. Regulation of metabolic pathways in liver and kidney during experimental diabetes: effects of antidiabetic compounds. Indian J Clin Bioche. 1998, 13(2): 63–80.

49] Saravanan G, Ponmurugan P, Senthil Kumar GP, Rajarajan T. Modulatory effect of S-allylcysteine on glucose metabolism in streptozotocin induced diabetes rats. J Func. Foods. 2009, 1: 336-40.

50] Vuppalapati L, Velayudam R, Ahamed KFHN, Cherukuri S, Kesavan R. The protective effect of dietary flavonoid fraction from Acanthophora spicifera on streptozotocin induced oxidative stress in diabetic rats. Food Sci Hum Well 2016, 5:57-64.

51] Goth L, Vitai M. Hypocatalasemia in hospital patients. Clin Chem 1996, 42:341-2.